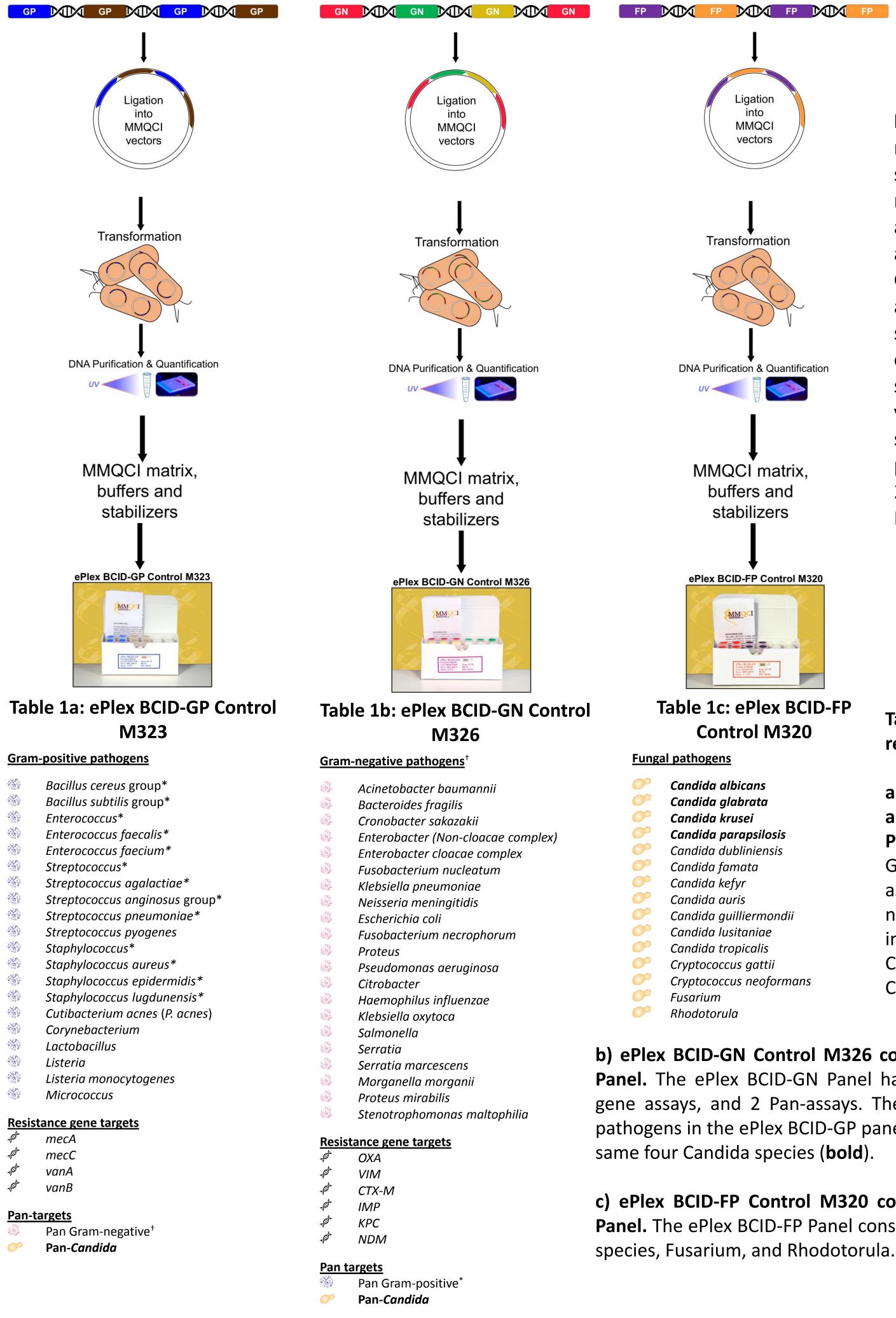


In-silico design

Bacterial and fungal blood infections that promote a hyperimmune response cause severe symptoms, including fever, pain, and tissue damage. If not treated quickly, this immune response can lead to a life-threatening condition called sepsis. Traditional blood culture methods including sub-culture isolations and antibiotic susceptibility testing take several days to obtain results. Rapid identification of bacterial pathogens and possible corresponding antibiotic resistance genes are critical to begin targeted treatment early. Reduced treatment time drastically diminishes the chance of lethality in septic patients.

Instruments that perform qualitative laboratory nucleic acid testing, such as GenMark's ePlex[®] system, aid in alleviating this issue; however, like any test in clinical use, their performance must be closely monitored to identify shifts, trends, and random errors caused by variations in the test system, such as failing reagents or operator errors. Maine Molecular Quality Controls Inc. (MMQCI) has developed a unique, extractable multiplex control panel designed to monitor all pathogenic organisms and antibiotic resistance genes detected by GenMark's ePlex[®] Blood Culture Identification (BCID) Gram-positive (GP), Gram-negative (GN), and Fungal Pathogen (FP) Panels

In-silico desigr



Session III (#41) Development of Synthetic Multiplexed External Controls for Monitoring the Performance of Qualitative Laboratory Nucleic Acid Testing Panels Used for Rapid Identification of Blood Culture Pathogens

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Materials and Methods

Figure 1: The synthetic, multiplex molecular controls contain genome segments of all Gram-positive, Gramnegative, and fungal pathogens, antimicrobial resistance markers, and Pan-targets that detected by the GenMark ePlex[®] BCID-GP, BCID-GN, BCID-FP Panels. Genome and segments were designed in silico to create several single pieces of synthetic DNA, ligated into MMQCI vectors, and transformed to create stable frozen clone stocks. DNA plasmids were purified, quantified by 260/280 UV spec, and formulated in MMQCI's proprietary matrix.

Table 1: ePlex BCID pathogens and resistance assays

a) ePlex BCID-GP Control M323 covers all 26 assays in the ePlex BCID-GP Panel. The ePlex BCID-GP Panel has 20 Gram-positive assays, 4 resistance gene assays, and 2 Pan-assays. The Pan Gramnegative assay covers all GN pathogens in the ePlex BCID-GN panel⁺. The Pan-Candida assay covers four common Candida species (**bold**).

b) ePlex BCID-GN Control M326 covers all 29 assays in the ePlex BCID-GN Panel. The ePlex BCID-GN Panel has 21 Gram-negative assays, 6 resistance gene assays, and 2 Pan-assays. The Pan Gram-positive assay covers 13 GP pathogens in the ePlex BCID-GP panel*. The Pan-Candida assay also covers the

c) ePlex BCID-FP Control M320 covers all 15 assays in the ePlex BCID-FP **Panel.** The ePlex BCID-FP Panel consists of 11 Candida species, 2 Cryptococcus

	1	able 2a: e	ePlex BCID	-GP Control M	A323 External Testing at GenMar	K			
ePlex BCID-GP Positive A (M324)	No. Samples Tested	No. Valid Tests	No. Samples Detected	Percent Samples Detected	ePlex BCID-GP Positive B (M325)	No. Samples Tested	No. Valid Tests	No. Samples Detected	Percent Samples Detected
<i>Bacillus cereus</i> group	79	77	77	100%	Cutibacterium acnes (P. acnes)	43	42	42	100%
Bacillus subtilis group	79	77	77	100%	Lactobacillus	43	42	42	100%
Corynebacterium	79	77	77	100%	Listeria	43	42	42	100%
Enterococcus	79	77	77	100%	Listeria monocytogenes	43	42	42	100%
Enterococcus faecalis	79	77	77	100%	Micrococcus	43	42	42	100%
Enterococcus faecium	79	77	77	100%	Staphylococcus	43	42	42	100%
Streptococcus	79	77	77	100%	Staphylococcus aureus	43	42	42	100%
Streptococcus agalactiae	79	77	77	100%	Staphylococcus epidermidis	43	42	42	100%
Streptococcus anginosus group	79	77	77	100%	Staphylococcus lugdunensis	43	42	42	100%
Streptococcus pneumoniae	79	77	77	100%	Pan <i>Candida</i>	43	42	42	100%
Streptococcus pyogenes	79	77	77	100%	Pan Gram-Negative	43	42	41	98%
vanA	79	77	77	77 100% mecA		43	42	42	100%
vanB	79	77	77	100%	mecC	43	42	42	100%
		Table 2b:	ePlex BCI	O-GP Control	M323 Internal Testing at MMQCI				
ePlex BCID-GP Positive A (M324)	No. Samples Tested	No. Valid Tests	No. Samples Detected	Percent Samples Detected	ePlex BCID-GP Positive B (M325)	No. Samples Tested	No. Valid Tests	No. Samples Detected	Percent Samples Detected
Bacillus cereus group	42	38*	38	100%	Cutibacterium acnes (P. acnes)	42	39	39	100%
Bacillus subtilis group	42	38*	37	97%	Lactobacillus	42	39	39	100%
Corynebacterium	42	39	39	100%	Listeria	42	39	39	100%
Enterococcus	42	39	39	100%	Listeria monocytogenes	42	39	39	100%
Enterococcus faecalis	42	39	39	100%	Micrococcus	42	39	39	100%
Enterococcus faecium	42	39	39	100%	Staphylococcus	42	39	39	100%
Streptococcus	42	39	39	100%	Staphylococcus aureus	42	39	39	100%
Streptococcus agalactiae	42	39	39	100%	Staphylococcus epidermidis	42	39	39	100%
Streptococcus anginosus group	42	39	39	100%	Staphylococcus lugdunensis	42	39	39	100%
Streptococcus pneumoniae	42	39	39	100%	Pan <i>Candida</i>	42	39	39	100%
Streptococcus pyogenes	42	39	39	100%	Pan Gram-Negative	42	39	39	100%
vanA	42	39	39	100%	mecA	42	38*	38	100%
vanB	42	39	39	100%	mecC	42	39	39	100%
Table 2: Verification of ePlex BC			1		ePlex Negative (M502)	No. Samples Tested	No. Valid Tests	No. Samples Not Detected	Percent Samples
					No targets	36	32	32	100%

No targets 36 32 32 100% GenMark (a) and MMQCI (b). a) MMQCI's ePlex BCID-GP Control M323 was tested a total of 122 times using 5 ePlex cartridge lots and 2 MMQCI control lots. Testing demonstrated robust and accurate results with 100% concordance across all Gram-positive targets and resistance gene targets in each control tubes. The Pan Gram-negative assay showed 98% concordance (42/43 runs detected). b) MMQCI performed a total of 84 runs across both positive controls and 40 negative control runs. Concordance of 100% was seen across 25/26 targets. *We removed 1 valid run from analysis that had cartridge-related performance issues causing false negatives in our control panel for mecA. B. subtilis, and B. cereus.

our control parter for meca	, D. Sustins, a								
		Table 3a:	ePlex BCID-	GN Control M	326 External Testing at GenN	Mark			
ePlex BCID-GN Positive A (M327)	No. Valid Tests		No. Samples Tested	No. Valid Tests	No. Samples Detected	Percent Samples Detected			
Acinetobacter baumannii	100	82	82	100%	Escherichia coli	100	91	90	99%
Bacteroides fragilis	100	82	82	100%	Fusobacterium necrophorum	100	91	88**	97%
Cronobacter sakazakii	100	82	82	100%	Proteus (spp.)	100	91	91	100%
<i>Enterobacter (</i> Non-cloacae complex <i>)</i>	100	82	82	100%	Pseudomonas aeruginosa	100	91	91	100%
<i>Enterobacter cloacae</i> complex	100	82	82	100%	Pan <i>Candida</i>	100	91	91	100%
Fusobacterium nucleatum	100	82	82	100%	Pan Gram-positive	100	91	91	100%
Klebsiella pneumoniae	100	82	82	100%	CTX-M	100	91	91	100%
Neisseria meningitidis	100	82	82	100%	ePlex BCID-GN Positive C (M329)	No. Samples Tested	No. Valid Tests	No. Samples Detected	Percent Samples Detected
Salmonella	100	82	82	100%	Citrobacter	98	91	91	100%
Serratia (spp.)	100	82	82	100%	Haemophilus influenzae	98	91	91	100%
Serratia marcescens	100	82	82	100%	Klebsiella oxytoca	98	91	91	100%
ΟΧΑ	100	82	82	100%	Morganella morganii	98	91	91	100%
VIM	100	82	82	100%	Proteus mirabilis	98	91	91	100%
					Stenotrophomonas maltophilia	98	91	91	100%
					IMP	98	91	91	100%
					КРС	98	91	91	100%

NDM

					KPC	98	91	91	100%
					NDM	98	91	91	100%
		Table 3b	: ePlex BCID	-GN Control N	1326 Internal Testing at MM	QCI			
ePlex BCID-GN Positive A (M327)	No. Samples Tested	No. Valid Tests	No. Samples Detected	Percent Samples Detected	ePlex BCID-GN Positive B (M328)	No. Samples Tested	No. Valid Tests	No. Samples Detected	Percent Samples Detected
Acinetobacter baumannii	11	10	10	100%	Escherichia coli	12	10	10	100%
Bacteroides fragilis	11	10	10	100%	Fusobacterium necrophorum	12	10	10	100%
Cronobacter sakazakii	11	10	10	100%	Proteus (spp.)	12	10	10	100%
<i>Enterobacter (</i> Non-cloacae complex <i>)</i>	11	10	10	100%	Pseudomonas aeruginosa	12	10	10	100%
Enterobacter cloacae complex	11	10	10	100%	Pan <i>Candida</i>	12	10	10	100%
Fusobacterium nucleatum	11	10	10	100%	Pan Gram-positive	12	10	10	100%
Klebsiella pneumoniae	11	10	10	100%	CTX-M	12	10	10	100%
Neisseria meningitidis	11	10	10	100%	ePlex BCID-GN Positive C (M329)	No. Samples Tested	No. Valid Tests	No. Samples Detected	Percent Samples Detected
Salmonella	11	10	10	100%	Citrobacter	10	10	10	100%
Serratia (spp.)	11	10	10	100%	Haemophilus influenzae	10	10	10	100%
Serratia marcescens	11	10	10	100%	Klebsiella oxytoca	10	10	10	100%
OXA	11	10	10	100%	Morganella morganii	10	10	10	100%
VIM	11	10	10	100%	Proteus mirabilis	10	10	10	100%
ePlex BCID-GN Negative (M503)	No. Samples Tested	No. Valid Tests	No. Samples Not Detected	Percent Samples Not Detected	Stenotrophomonas maltophilia	10	10	10	100%
No targets	13	12	12	100%	IMP	10	10	10	100%

Table 3: Verification of ePlex BCID-GN Control M326 at GenMark (a) and MMQCI (b). a) MMQCI's ePlex BCID-GN Control tested was highly reproducible and robust with 100% **Discordant calls for F. necrophorum were due to **b)** MMQCI performed a total of 46 runs across all pos

Results

I M326 was tested a total	of 298 times using 12 ePlex cart	ridge lots and 3 MMQ	CI control lots. M326
% concordance across 19/2	21 Gram-negative targets and re	esistance gene targets i	n each control tube.
o cartridge-related perfo	rmance issues. E. coli showed	99% concordance (90	/91 runs detected).
sitive and negative contro	and achieved 100% concordan	ice.	

Candida albicar Candida famata Candida glabrata Candida kefyr Cryptococcus gatti

Candida dubilnien Candida famata Candida glabrate Candida kefyr Cryptococcus gatti

cartridge.

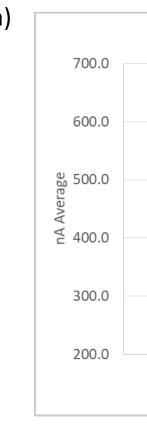


Figure 2: Representative Real-time Functional Stability of ePlex BCID Products a) BCID-GP Positive A (M324) was tested for realtime functional stability on the ePlex system over the course of 14 months. The graphs shows average nanoamp (nA) values across all targets, which is the signal generated by the ePlex instrument. b) BCID-GP Positive B (M325) was tested for real-time functional stability on the ePlex system over the course of 10 months. The graphs shows average nanoamp (nA) values across all targets, which is the signal generated by the ePlex instrument. Linear regression analysis shows no significant trending with significance of *F value* > 0.05. Testing is on-going

MM	Q
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100%

10

This study was performed using only GenMark RUO consumables. ePlex[®] In-vitro Diagnostic (IVD) material is expected to have higher validity upon launch.

MMQCI's proprietary matrix and stabilization buffers allow for stable, reliable controls that can be carried through the entire molecular diagnostic assay to accurately simulate all pathogens and corresponding resistance markers detected by GenMark's ePlex[®] Blood Culture Identification (BCID) Panels assay.

Controls performed robustly at both sites with >97% accuracy across all targets for all three panels.

MMQCI's ePlex BCID Controls are ready-to-use, non-infectious and well-characterized quality control panels for use in the clinical laboratory.

Current real time stability data indicate stability over 12 months, based on data from products with similar formulation, we predict stability of 24-months for all ePlex BCID controls.

			-							
	Table 4a: ePlex BCID-FP Control M320 External Testing at GenMark									
ositive A	No. Samples	No. Valid	No. Samples	Percent Samples	ePlex BCID-FP Positive B	No. Samples	No. Valid	No. Samples	Percent Samples	
	Tested	Tests	Detected	Detected	(M322)	Tested	Tests	Detected	Detected	
	97	94	94	100%	Candida auris	125	122	122	100%	
sis	97	94	94	100%	Candida guilliermondii	125	122	122	100%	
	97	94	94	100%	Candida krusei	125	122	122	100%	
	97	94	94	100%	Candida lusitaniae	125	122	122	100%	
	97	94	94	100%	Candida parapsilosis	125	122	122	100%	
i	97	94	94	100%	Candida tropicalis	125	122	122	100%	
					Cryptococcus neoformans	125	122	122	100%	
					Fusarium	125	122	122	100%	
					Rhodotorula	125	122	120 [‡]	98%	

Table 4b: ePlex BCID-FP Control M320 Internal Testing at MMQCI

ositive A	No. Samples Tested	No. Valid Tests	No. Samples Detected	Percent Samples Detected
	13	12	12	100%
sis	13	12	12	100%
	13	12	12	100%
	13	12	12	100%
	13	12	12	100%
ii	13	12	12	100%

Table 4: Verification of ePlex BCID-FP Control M320 at GenMark (a) and MMQCI (b). a) The ePlex BCID-FP Control M320 was tested 222 times at GenMark and

(M322)	Tested	Tests	Detected	Detected
Candida auris	34	33*	33	100%
Candida guilliermondii	34	34	34	100%
Candida krusei	34	34	34	100%
Candida lusitaniae	34	34	34	100%
Candida parapsilosis	34	34	34	100%
Candida tropicalis	34	34	34	100%
Cryptococcus neoformans	34	34	34	100%
usarium	34	34	34	100%
Rhodotorula	34	33*	32	97%

achieved 100% concordance across 14/15 targets. [‡] The 2 false negatives seen for Rhodotorula occurred on the same cartridge lot; 1 false negative was due to an issue with an amplification pool. b) M320 testing at MMQCI showed 100% concordance across 14/15 targets. *We observed 1 valid run that had multiple false negatives due to a bay/cartridge problem that was excluded from our analyses. The 1 Rhodotorula false negative was due to poor PCR pool amplification on that particular

M324 eF	Plex BCID-GP Po Real Time S	ositive A (E06FEB Stability	18)	b)	700.0	M325 ePlex	BCID-GP Positive B Real Time Stability	(C08JUN18)
					700.0 600.0 80 500.0 90 400.0			
					200.0			
	6 Mor	12 aths	14		200.0	0	2 Months	10

Conclusions

Cl's synthetic, multiplex controls are designed to be part of an essential clinical laboratory control program. The multiplex nature of the controls streamlines manufacture, thus g them affordable for clinical labs.

Additional validation data resulted in a post-launch product improvement to M323.

Acknowledgements