

Background

Pneumonia is an infection of the lungs that results in hospitalization of over a million Americans per year in the United States. Classical microbiological methods take days to obtain results. For patients hospitalized with pneumonia, a fast diagnosis is critical to prevent mortality. Panel-based molecular diagnostic assays that can identify multiple pathogens and antibiotic resistance are newly available to aid in the diagnosis of lower respiratory infections and significantly shorten the time to effective targeted therapy. Performance of all clinical assays must be closely monitored to identify shifts, trends, and random errors in order to ensure accurate results. A multiplex control panel has been designed to monitor all analytes detected by the FDA-cleared Curetis Unyvero Lower Respiratory Tract (LRT) Application.



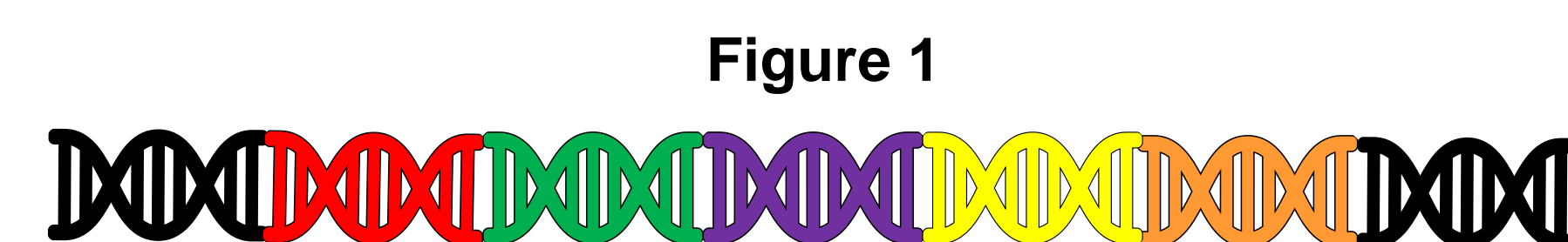
Multiplexed quality controls help with:

- Detecting system errors
- Identifying shifts/trends of data over time
- Equal QC representation of all assay targets, not just easy-to-obtain pathogen samples
- Fast and efficient monitoring of multiplex assays
- Ease-of-use: multiple targets in just a few external controls allows for easier tracking

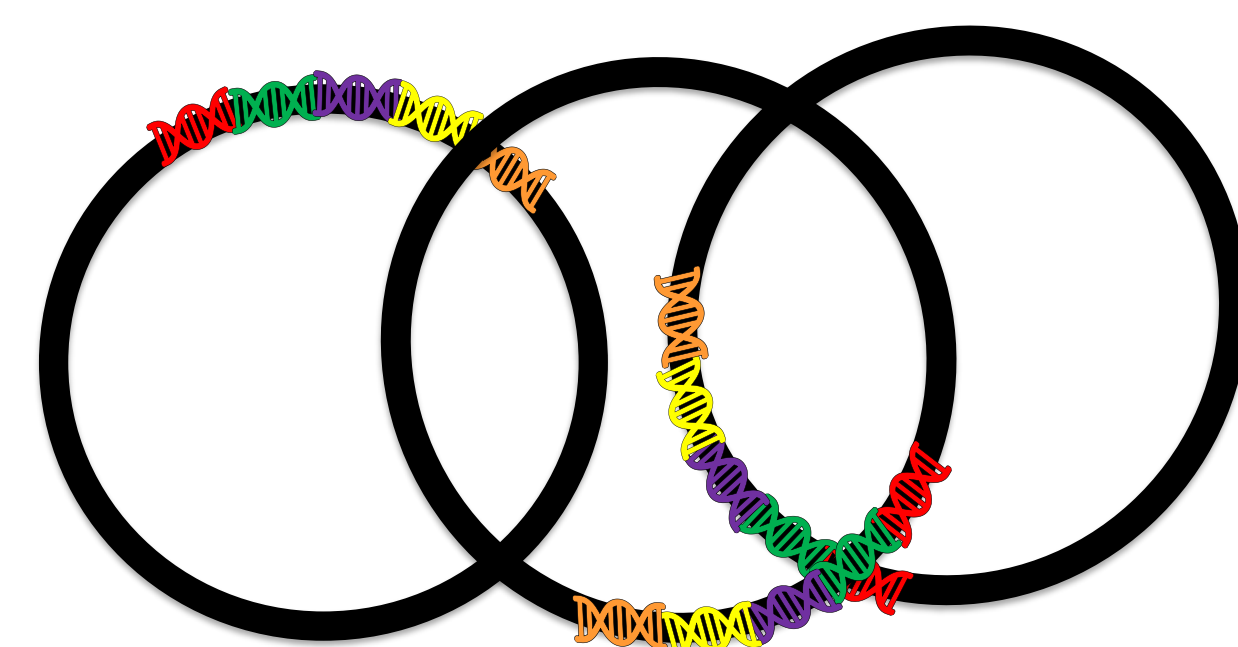
Materials and Methods

The synthetic, multiplex molecular controls contain genome segments of all 19 bacterial pathogens and their corresponding 10 resistance markers detected by the FDA-cleared Curetis Unyvero LRT Application. Pathogen segments were designed in silico, dispersed among several large pieces of synthetic DNA, ligated into engineered vectors, and transformed to create stable frozen clone stocks. DNA plasmids were purified, quantified by 260/280 UV spec, and formulated in a proprietary matrix which carries the control DNA first through a rigorous lysis process and ending in a sensitive array detection system (Figure 1).

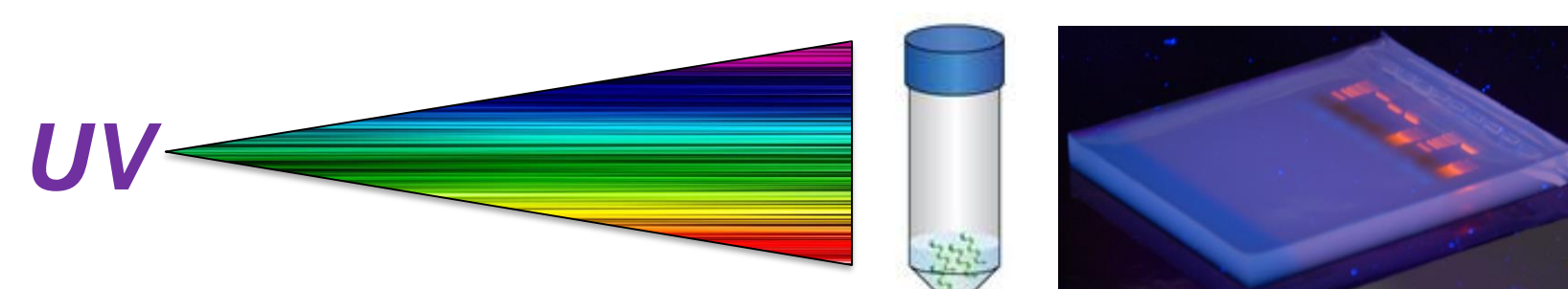
1) *In silico* DNA synthesis of genomic segments from pathogens detected by the Unyvero LRT Application



2) Sub-cloning onto MMQCI engineered vectors



3) Plasmid Purification/Quantification



4) MMQCI Formulation (Matrix and stabilizers)



Table 1: Unyvero LRT Application pathogens and resistance assays.

Unyvero LRT/Pneumonia POS A (M405)	Unyvero Microorganism Classification
<i>Enterobacter cloacae</i> complex	Enterobacteriaceae (Gram-negative)
<i>Escherichia coli</i>	Enterobacteriaceae (Gram-negative)
<i>Klebsiella oxytoca</i>	Enterobacteriaceae (Gram-negative)
<i>Klebsiella pneumoniae</i>	Enterobacteriaceae (Gram-negative)
<i>Klebsiella variicola</i>	Enterobacteriaceae (Gram-negative)
<i>Serratia marcescens</i>	Enterobacteriaceae (Gram-negative)
<i>Staphylococcus aureus</i>	Gram-positive
<i>Pseudomonas aeruginosa</i>	Non-fermenting
<i>Mycoplasma pneumoniae</i>	Other
ctx-M	Antibiotic resistance marker (3rd Gen. Cephalosporins)
kpc	Antibiotic resistance marker (Carbapenems)
ndm	Antibiotic resistance marker (Carbapenems)
oxa-48	Antibiotic resistance marker (Carbapenems)
vim	Antibiotic resistance marker (Carbapenems)
Unyvero LRT/Pneumonia POS B (M406)	Unyvero Microorganism Classification
<i>Citrobacter freundii</i>	Enterobacteriaceae (Gram-negative)
<i>Morganella morganii</i>	Enterobacteriaceae (Gram-negative)
<i>Proteus</i> spp.	Enterobacteriaceae (Gram-negative)
<i>Staphylococcus aureus</i>	Gram-positive
<i>Streptococcus pneumoniae</i>	Gram-positive
<i>Acinetobacter</i> spp.	Non-fermenting
<i>Stenotrophomonas maltophilia</i>	Non-fermenting
<i>Chlamydia pneumoniae</i>	Other
<i>Haemophilus influenzae</i>	Other
<i>Legionella pneumophila</i>	Other
<i>Moraxella catarrhalis</i>	Other
mecA	Antibiotic resistance marker (Oxacillin/Cefoxitin)
oxa-24	Antibiotic resistance marker (Carbapenems)
oxa-23	Antibiotic resistance marker (Carbapenems)
oxa-58	Antibiotic resistance marker (Carbapenems)
tem	Antibiotic resistance marker (Penicillins)
Unyvero LRT/Pneumonia NEG (M407)	Unyvero Microorganism Classification
No targets present	N/A

Unyvero LRT/Pneumonia NEG (M407)	No. samples tested	No. valid tests	No. samples not detected	Percent samples not detected
<i>Enterobacter cloacae</i> complex	38	38	38	100%
<i>Escherichia coli</i>	38	38	38	100%
<i>Klebsiella oxytoca</i>	38	38	38	100%
<i>Klebsiella pneumoniae</i>	38	38	38	100%
<i>Klebsiella variicola</i>	38	38	38	100%
<i>Serratia marcescens</i>	38	38	38	100%
<i>Staphylococcus aureus</i>	38	38	38	100%
<i>Pseudomonas aeruginosa</i>	38	38	38	100%
<i>Mycoplasma pneumoniae</i>	38	38	38	100%
ctx-M	38	38	38	100%
kpc	38	38	38	100%
ndm	38	38	38	100%
oxa-48	38	38	38	100%
vim	38	38	38	100%
<i>Citrobacter freundii</i>	38	38	38	100%
<i>Morganella morganii</i>	38	38	38	100%
<i>Proteus</i> spp.	38	38	38	100%
<i>Staphylococcus aureus</i>	38	38	38	100%
<i>Streptococcus pneumoniae</i>	38	38	38	100%
<i>Acinetobacter</i> spp.	38	38	38	100%
<i>Stenotrophomonas maltophilia</i>	38	38	38	100%
<i>Chlamydia pneumoniae</i>	38	38	38	100%
<i>Haemophilus influenzae</i>	38	38	38	100%
<i>Legionella pneumophila</i>	38	38	38	100%
<i>Moraxella catarrhalis</i>	38	38	38	100%
mecA	38	38	38	100%
oxa-24	38	38	38	100%
oxa-23	38	38	38	100%
oxa-58	38	38	38	100%
tem	38	38	38	100%

Table 2 continued. M407 NEG: No targets were detected in 38/38 negative control runs on the Unyvero A50 System (100% concordant).

Results

Table 2: Verification of Unyvero LRT/Pneumonia Control Panel M404

Unyvero LRT/Pneumonia POS A (M405)	No. samples tested	No. valid tests	No. samples detected	Percent samples detected
<i>Enterobacter cloacae</i> complex	37	31	31	100%
<i>Escherichia coli</i>	37	31	31	100%
<i>Klebsiella oxytoca</i>	37	31	30	96.8%
<i>Klebsiella pneumoniae</i>	37	31	31	100%
<i>Klebsiella variicola</i>	37	31	31	100%
<i>Serratia marcescens</i>	37	31	31	100%
<i>Staphylococcus aureus</i>	37	31	30	96.8%
<i>Pseudomonas aeruginosa</i>	37	31	31	100%
<i>Mycoplasma pneumoniae</i>	37	31	31	100%
ctx-M	37	31	31	100%
kpc	37	31	31	100%
ndm	37	31	31	100%
oxa-48	37	31	31	100%
vim	37	31	31	100%
Unyvero LRT/Pneumonia POS B (M406)	No. samples tested	No. valid tests	No. samples detected	Percent samples detected
<i>Citrobacter freundii</i>	44	44	44	100%
<i>Morganella morganii</i>	44	44	44	100%
<i>Proteus</i> spp.	44	44	44	100%
<i>Staphylococcus aureus</i>	44	44	44	100%
<i>Streptococcus pneumoniae</i>	44	44	43	97.7%
<i>Acinetobacter</i> spp.	44	44	44	100%
<i>Stenotrophomonas maltophilia</i>	44	44	44	100%
<i>Chlamydia pneumoniae</i>	44	44	44	100%
<i>Haemophilus influenzae</i>	44	44	44	100%
<i>Legionella pneumophila</i>	44	44	44	100%
<i>Moraxella catarrhalis</i>	44	44	44	100%
mecA	44	44	44	100%
oxa-24	44	44	44	100%
oxa-23	44	44	44	100%
oxa-58	44	44	44	100%
tem	44	44	44	100%

Table 2. M405 POS A: Overall, 30/31 runs showed 100% concordance on the Unyvero A50 System. Six runs were removed from analysis due to a manufacturing defect in 1 particular cartridge lot. One run showed false negatives for PCR pool 6 assays *K. oxytoca* and *S. aureus* due to chamber 6 internal control failure. M406 POS B: Overall, 43/44 runs showed 100% concordance. One run showed a false negative for PCR pool 6 assay *S. pneumoniae* due to chamber 6 internal control failure.

Figure 2a

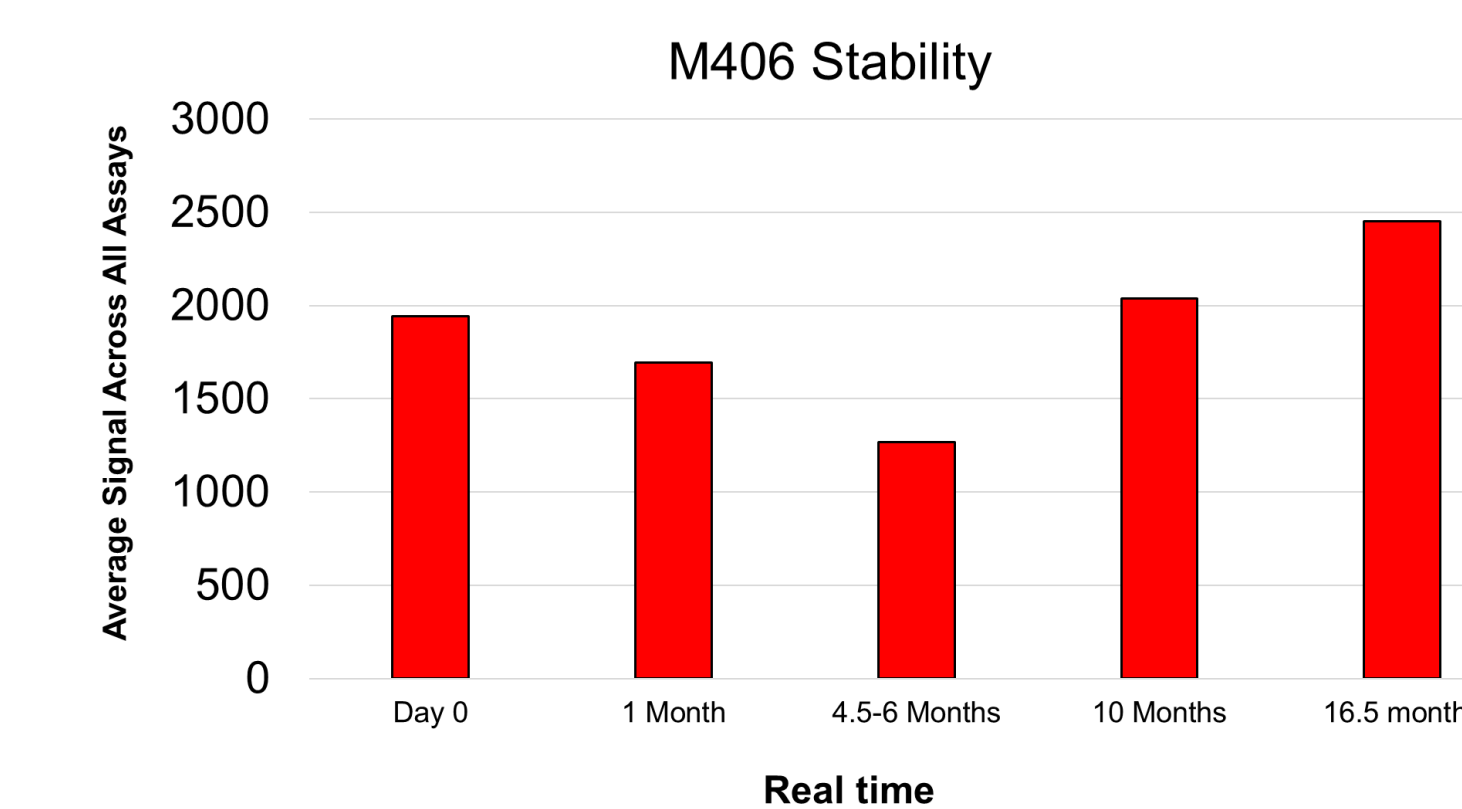


Figure 2b

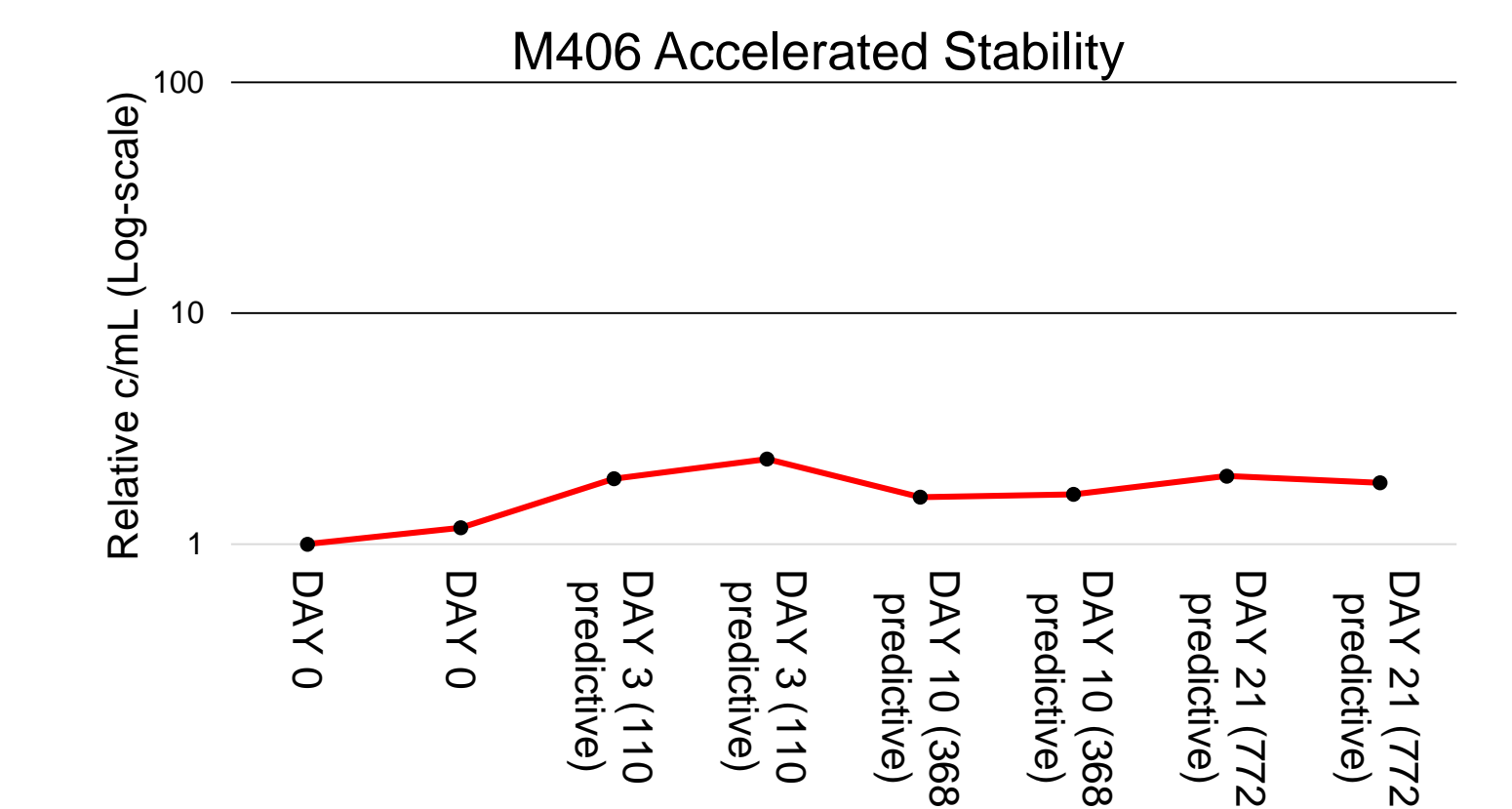


Figure 2: Representative Real-time Stability and Accelerated stability of Unyvero LRT/Pneumonia POS B (M406) a) One control lot of M406 was tested on the Unyvero A50 system over the course of 16.5 months. Signal across all LRT assays were averaged together for the specific time point. Replicates: Day 0 (n=10), 1 month (n=6), 4.5-6 months (n=10), 10 months (n=5), 16.5 months (n=4). b) One control lot was placed at 56°C for 3-21 days to assess accelerated stability. Samples were extracted using the QIAmp DNA Blood kit. An in-house MMQCI qPCR assay was run on the Roche Light Cycler to determine c/mL. Relative c/mL from day 0 was graphed on a log-scale. Using the Arrhenius temperature coefficient equation with a Q10 value of 2, this would be predictive of a ~2.1 years stability when stored at 2-8°C.

Conclusions

- MMQCI's synthetic, multiplex controls are designed to be part of an essential clinical laboratory quality control program. The multiplex nature of the controls streamlines the laboratory's Individualized Quality Control Plan (IQCP) without decreasing stringency.
- MMQCI's proprietary matrix and stabilization buffers allow for stable, reliable controls that can be carried through the Unyvero mechanical/chemical lysis, cartridge purification, PCR, and array detection.
- Controls performed robustly with ≥96% accuracy across all targets on the Unyvero LRT application.
- MMQCI's Curetis Unyvero controls are ready-to-use, non-infectious and well-characterized quality control panels for use in the clinical laboratory.
- Current accelerated stability data indicates stability of 24-months. On-going real-time stability demonstrates stability of over 12 months

Acknowledgements: Reagents for this study provided by Curetis USA, San Diego, CA and Curetis (Germany)